

TakaRa

RetroNectin™

in Gene Therapy Related Studies



BIBLIOGRAPHY

Characteristics and Properties of RetroNectin™

RetroNectin™ (CH-296) is a chimeric peptide of recombinant human fibronectin fragments. Takara's proprietary expression system constructed in *E. coli* efficiently generates fragment variations as shown in Fig. 1: Fibronectins (FNs) are multi-functional cell adhesive glycoproteins present in

extracellular matrix and plasma. RetroNectin™ is comprised of 574 amino acids (63 kDa) and has three functional domains; central cell-binding domain (type III repeat, 8, 9, 10), heparin-binding domain II (type III repeat, 12, 13, 14), and CS-1 site within the alternatively spliced IIICS region.

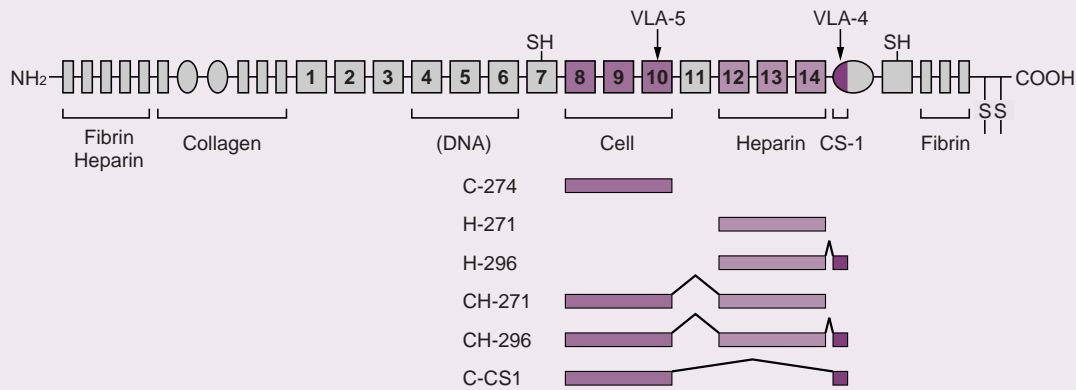


Fig. 1. Structure of FN and Fragments including RetroNectin™

Kimizuka F, et al. Production and Characterization of Functional Domains of Human Fibronectin Expressed in *Escherichia coli*. *J. Biochem.* 110, 284-291 (1991)

When coated on the surface of any containers such as culture dishes, well plates, flasks or bags, RetroNectin™ significantly enhances retrovirus-mediated gene transfer into mammalian cells. This enhancement is hypothetically attributed to the colocalization of retroviral particles and target cells on the molecules of RetroNectin™ (Fig. 2). Virus particles bind to this molecule

through the interaction with the heparin-binding domain II. Target cells can be localized on RetroNectin™ mainly through the interaction of the fibronectin CS-1 site with very late antigen 4 (VLA-4) and/or through the RGDS sequence in repeat 10 of fibronectin Cell-binding domain with very late antigen 5 (VLA-5).

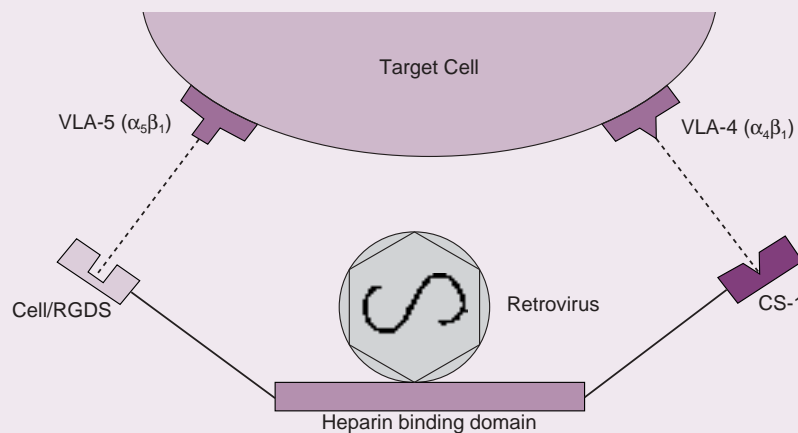


Fig. 2. Hypothetical Mechanism of Transduction Enhancement by RetroNectin™ (Recombinant Human Fibronectin Fragment CH-296)

Hanenberg H, et al. Colocalization of Retrovirus and Target Cells on Specific Fibronectin Fragments Increases Genetic Transduction of Mammalian Cells. *Nature Medicine* 2(8), 876-882 (1996)

Meeting Challenges Toward the Gene Therapy Reality in the 21st Century

TAKARA BIO INC. has contributed greatly to the progress in gene transfer and related fields including hematopoietic cell gene therapy by supplying a recombinant human fibronectin fragment CH-296, or RetroNectin™ for over half a decade. RetroNectin™ is a key component in successful clinical protocols for retroviral-mediated cell transduction in terms of safety and efficiency.

Notes:

*1 FN CH-296 for Clinical Trial Use is produced at GMP facilities in compliance with related regulations, while commercially available RetroNectin™ for Research Use Only is manufactured at completely separate facilities.

*2 RetroNectin™ related patents TAKARA BIO holds:
Patents Issued: US Patents No. 5198423, No. 5686278 & No. 6033907 and European Patent No. 399806
Patents Pending: US Patent Applications No. 09/043,981 & No. 09/394,867 and European Patent Applications No. 95914912.1 & No. 96933209.7

The year 2000 was a watershed in the gene therapy field. In addition to the successful outcomes of the human X-SCID gene therapy in France, there have been conspicuous achievements in clinical trials in several other countries. The latest trials that have been treating congenital diseases were presented at the 5th Annual Meeting of the American Society of Gene Therapy (ASGT) as interim reports of ongoing trials.

Aiuti A, Slavin S, Aker M, et al. Correction of ADA-SCID by Stem Cell Gene Therapy Combined with Nonmyeloablative Conditioning. *Science* 296, 2410-2413 (2002)

Hacein-Bey-Abina S, Le Deist F, Carlier F, et al. Sustained Correction of X-linked Severe Combined Immunodeficiency by *Ex Vivo* Gene Therapy. *N Engl J Med* 346(16), 1185-1193 (2002)

Cavazzana-Calvo M, Hacein-Bey S, Basile GS, et al. Gene Therapy of Human Severe Combined Immunodeficiency (SCID)-X1 Disease. *Science* 288, 669-672 (2000)

Abonour R, Williams DA, Einhorn L, et al. Efficient Retrovirus-Mediated Transfer of the Multidrug Resistance 1 Gene into Autologous Human Long-Term Repopulating Hematopoietic Stem Cells. *Nature Medicine* 6(6), 652-658(2000)

Malech HL. Use of Serum-Free Medium with Fibronectin Fragment Enhanced Transduction in a System of Gas Permeable Plastic Containers to Achieve High Levels of Retrovirus Transduction at Clinical Scale. *Stem Cells* 18, 155-156 (2000)

Smaglik P. For Stem Cell Transduction, The Solution Is in the Bag. *The Scientist* 13(15), July 19 (1999)

Bauer G, Selander D, Engel B, et al. Gene Therapy for Pediatric AIDS. *Annals New York Academy of Science* 918, 318-329 (2000)

Schmidt M, Hacein-Bey S, LeDeist F, et al. Detection of Pluripotent Hematopoietic Cells in SCID-X1 Gene Therapy. *Mol Ther* 5(5), Abstract 68, p. S25 (2002)

Parsley K, Gilmour K, Brouns G, et al. Successful Treatment of Human X-SCID Using a GALV-Pseudotyped Retroviral Vector. *Mol Ther* 5(5), Abstract 70, p. S26 (2002)

Candotti F, Podsakoff GM, Schurman SH, et al. Evaluation of Adenosine Deaminase (ADA) Gene Transfer by Retroviral Vectors for the Treatment of SCID. *Mol Ther* 5(5), Abstract 888, p. S289 (2002)

Aiuti A, Slavin S, Aker M, et al. Correction of ADA-SCID by Stem Cell Gene Therapy Combined with a Non-Myeloablative Conditioning. *Mol Ther* 5(5), Abstract 931, p. S304 (2002)

There have been several comprehensive review articles published that summarize the past progress and recent advances in strategies for efficient and optimized gene transfer into hematopoietic cells. Several also foresee a wide range of applications to be realized as the choice of therapy in the 21st century. However, there will be many problems to be addressed and overcome.

Anderson WF. The Current Status of Clinical Gene Therapy. *Hum Gene Ther* 13, 1261-1262 (2002) [Editorial]

Bordignon C, Roncarolo MG. Therapeutic Applications for Hematopoietic Stem Cell Gene Transfer. *Nature Immunol* 3(4), 318-321 (2002) [Comment]

Candotti F. Gene Therapy for Immunodeficiency. *Curr Allergy Asthma Rep* 1(5), 407-415 (2001) [Review]

Thomas AK, June CH. The Promise of T-Lymphocyte Immunotherapy for the Treatment of Malignant Disease. *Cancer J* 7, Suppl 2, S67-S75 (2001) [Review]

Williams DA, Nienhaus AW, Hawley RG, Smith FO. Gene Therapy 2000. *Hematology 2000*, 376-393 (2000) [Review]

Anderson WF. Gene Therapy. The Best of Times, the Worst of Times. *Science* 288, 627-629 (2000) [Comment]

Dick JE. Gene Therapy Turns the Corner. *Nature Medicine* 6(6), 624-626 (2000) [Comment]

Halene S and Kohn DB. Gene Therapy Using Hematopoietic Stem Cells: Sisyphus Approaches the Crest. *Hum Gene Ther* 11, 1259-1267 (2000) [Review]

Fischer A. Cautious Advance: Gene Therapy is More Complex Than Anticipated. *EMBO Reports* 1(4), 284-296 (2000) [Review]

Williams DA and Smith FO. Progress in the Use of Gene Transfer Methods to Treat Genetic Blood Diseases. *Hum Gene Ther* 11, 2059-2066 (2000) [Review]

Improved transduction protocols and successful translation of preclinical methods, which have been developed and evaluated *in vivo* with animals from mice to canines to non-human primates, have enhanced human trial outcomes. Improvement in therapeutic strategies for gene transfer into hematopoietic cells continues to develop in order to modulate immune responses, to protect hematopoietic cells against cytotoxic drugs or viral genes, and to restore congenital or acquired gene deficiencies.

Cell transplantation following cancer treatment such as chemotherapy has been expanded to utilize gene therapy in order to introduce drug-resistance genes for protecting hematopoietic cells and the HSV-tk suicide gene for controlling GVHD in allogeneic transplantation that are widely practiced for the treatment of malignant diseases. A further use envisioned is immunogene therapy to increase the immunogenicity of tumor cells or cytotoxicity of specific cells to kill tumors. A variety of adoptive cellular immunotherapy strategies using *ex vivo* gene transfer have aimed at boosting the immune system; these *ex vivo* strategies include gene delivery into cellular components of the immune system, such as cytotoxic T cells, NK cells, macrophages and dendritic cells.

Qasim W, Thrasher AJ, Buddle J. et al. T Cell Transduction and Suicide with an Enhanced Mutant Thymidine Kinase. *Gene Ther* 9(12), 824-827 (2002)

Lamers CHJ, Willemsen RA, Luiders BA, et al. Protocol for Gene Transduction and Expansion of Human T Lymphocytes for Clinical Immunogene Therapy of Cancer. *Cancer Gene Ther* 9, 613-623 (2002)

Liu K, Rosenberg SA. Transduction of an *IL-2* Gene into Human Melanoma-Reactive Lymphocytes Results in Their Continued Growth in the Absence of Exogenous IL-2 and Maintenance of Specific Antitumor Activity. *J Immunol* 167, 6356-6365 (2001)

Ragg S, Xu-Welliver M, Bailey J, et al. Direct Reversal of DNA Damage by Mutant Methyltransferase Protein Protects Mice Against Dose-Intensified Chemotherapy and Leads to *In Vivo* Selection of Hematopoietic Stem Cells. *Cancer Res* 60, 5187-5195 (2000)

Schildmeier B, Wermann K, Kühlcke K, et al. Human Multidrug Resistance-1 Gene Transfer to Long-Term Repopulating Human Mobilized Peripheral Blood Progenitor Cells. *Bone marrow Transplant* 25, Suppl.2, S118-S124 (2000)

Schilz AJ, Schildmeier B, Kühlcke K, et al. *MDR1* Gene Expression in NOD/SCID Repopulating Cells after Retroviral Gene Transfer under Clinically Relevant Conditions. *Mol Ther* 2(6), 609-618 (2000)

Schildmeier B, Kühlcke K, Eckert HG, et al. Quantitative Assessment of Retroviral Transfer of the Human Multidrug Resistance 1 Gene to Human Mobilized Peripheral Blood Progenitor Cells Engrafted in Nonobese Diabetic/Severe Combined Immunodeficient Mice. *Blood* 95(4), 1237-1248 (2000)

Kuga T, et al. Fibronectin Fragment-Facilitated Retroviral Transfer of the Glutathione-S-Transferase p Gene into CD34+ Cells to Protect Them Against Alkylating Agents. *Hum Gene Ther* 8(16), 1901-1910 (1997)

Maze R, et al. Establishing Chemoresistance in Hematopoietic Progenitor Cells. *Mol Med Today* 3(8), 350-358 (1997)

Gene transfer into T-cells that can be facilitated with the use of FN CH-296 is another major advance in order to treat GVHD or AIDS. From the perspective of immuno-gene therapy, T-cell manipulation may also be incorporated in cancer gene therapy protocols.

Eaton D, Gilham DE, O'Neil A, Hawkins RE. Retroviral Transduction of Human Peripheral Blood Lymphocytes with Bcl-X(L) Promotes *In Vitro* Lymphocytes Survival in Pro-Apoptotic Conditions. *Gene Ther* 9(8), 527-535 (2002)

Koehne G, Gallardo HF, Sadelain M and O'Reilly RJ. Rapid Selection of Antigen-Specific T Lymphocytes by Retroviral Transduction. *Blood* 96(1), 109-117 (2002)

Zhou P, Lee J, Moore P, et al. High-Efficiency Gene Transfer into Rhesus Macaque Primary T Lymphocytes by Combining 32 °C Centrifugation and CH-296-Coated Plates : Effect of Gene Transfer Protocol on T Cell Homing Receptor Expression. *Hum Gene Ther* 12, 1843-1855 (2001)

Lamana ML, Segovia JC, Guenechea G, et al. Systematic Analysis of Clinically Applicable Conditions Leading to a High Efficiency of Transduction and Transgene Expression in Human T Cells. *J Gene Med* 3, 32-41 (2001)

Dardalhon V, Herpers B, Noraz N, et al. Lentivirus-Mediated Gene Transfer in Primary T Cells is Enhanced by a Central DNA Flap. *Gene Ther* 8, 190-198 (2001)

Fujio K, Misaki Y, Setoguchi K, et al. Functional Reconstitution of Class II MHC-Restricted T Cell Immunity Mediated by Retroviral Transfer of the $\alpha\beta$ TCR Complex. *J Immunol* 165, 528-532 (2000)

Dardalhon V, Jaleco S, Rebouissou C, et al. Highly Efficient Gene Transfer in Naive Human T Cells with a Murine Leukemia Virus-Based Vector. *Blood* 96(3), 885-893 (2000)

Movassagh M, Boyer O, Burland MC, et al. Retrovirus-Mediated Gene Transfer into T Cells: 95% Transduction Efficiency without Further *In Vitro* Selection. *Hum Gene Ther* 11(8), 1189-1200 (2000)

Koehne G, Gallardo HF, Sadelain M, et al. Rapid Selection of Antigen-Specific T Lymphocytes by Retroviral Transduction. *Blood* 96(1), 109-117 (2000)

Kühlcke K, Ayuk FA, Li Z, et al. Retroviral Transduction of T Lymphocytes for Suicide Gene Therapy in Allogeneic Stem Cell Transplantation. *Bone Marrow Transplant* 25(2), S96-S98 (2000)

Costello E, Munoz M, Buetti E, et al. Gene Transfer into Stimulated and Unstimulated T Lymphocytes by HIV-1-Derived Lentiviral Vectors. *Gene Ther* 7, 596-604 (2000)

Uckert W, Becker C, Gladow M, et al. Efficient Gene Transfer into Primary Human CD8+ T Lymphocytes by MuLV-10A1 Retrovirus Pseudotype. *Hum Gene Ther* 11, 1005-1014 (2000)

Steinberg M, Swainson L, Schwarz K, et al. Retrovirus-Mediated Transduction of Primary ZAP-70-Deficient Human T Cells Results in the Selective Growth Advantage of Gene-Corrected Cells: Implications for Gene Therapy. *Gene Ther* 7, 1392-1400 (2000)

RetroNectin™-assisted Gene Transfer Protocols that also improve biological safety and standardization have considerable potential in impacting on the use of gene therapy for a variety of diseases.

Otsu M, Steinberg M, Ferrand C, et al. Reconstitution of Lymphoid Development and Function in ZAP-70-Deficient Mice Following Gene Transfer into Bone Marrow Cells. *Blood* 100(4), 1248-1256 (2002)

Kurre P, Morris J, Andrews RG, et al. Kinetics of Fluorescence Expression in Nonhuman Primates Transplanted with GFP Retrovirus-Modified CD34 Cells. *Mol Ther* 6(1), 83-90 (2002)

Linderman C, Schilz AJ, Emons B, et al. Down-Regulation of Retroviral Transgene Expression during Differentiation of Progenitor-Derived Dendritic Cells. *Exp Hematol* 30(2), 150-157 (2002)

Kiem HP, Rasko JEJ, Morris J, et al. *Ex Vivo* Selection for Oncoretrovirally Transduced Green Fluorescent Protein-Expressing CD34-Enriched Cells Increases Short-Term Engraftment of Transduced Cells in Baboons. *Hum Gene Ther* 13,891-899 (2002)

Dao MA, Hwa J, Nolte JA, et al. Molecular Mechanism of Transforming Growth Factor β -Mediated Cell-Cycle Modulation in Primary Human CD34⁺ Progenitors. *Blood* 99(2), 499-506 (2002)

Tao W, Filippi MD, Bailey JR, et al. The TRQQRK Motif Located Near the C-Terminus of Rac2 is Essential for Rac2 Biologic Functions and Intracellular Localization. *Blood* 100(5), 1679-1688 (2002)

Weijtens M, Spronsen AV, Hagenbeek A, et al. Reduced Graft-Versus-Host Disease-Inducing Capacity T Cells After Activation, Culturing, and Magnetic Cell Sorting Selection in an Allogeneic Bone Marrow Transplantation Model in Rats. *Hum Gene Ther* 13, 187-198 (2002)

Van Der Loo J, Liu BL, Goldman AI, et al. Optimization of Gene Transfer into Primitive Human Hematopoietic Cells of Granulocyte-Colony Stimulating Factor-Mobilized Peripheral Blood Using Low-Dose Cytokines and Comparison of a Gibbon Ape Leukemia Virus Versus an RD114-Pseudotyped Retroviral Vector. *Hum Gene Ther* 13,1317-1330 (2002)

Björgvinsdóttir H, Bryder D, Sitnicka E, et al. Efficient Oncoretroviral Transduction of Extended Long-Term Culture-Initiating Cells and NOD/SCID Repopulating Cells: Enhanced Reconstitution with Gene-Marked Cells Through an *Ex Vivo* Expansion Approach. *Hum Gene Ther* 13, 1061-1073 (2002)

Wada T, Jagadeesh GJ, Nelson DL, et al. Retrovirus-Mediated WASP Gene Transfer Corrects Wiskott-Aldrich Syndrome T-Cell Dysfunction. *Hum Gene Ther* 13, 1039-1046 (2002)

Hanenberg H, Batish SD, Pollok KE, et al. Phenotypic Correction of Fanconi Anemia T Cells with Retroviral Vectors as a Diagnostic Tool. *Exp Hematol* 30, 410-420 (2002)

Bhatia R, Williams AD, Munthe HA, et al. Contact with Fibronectin Enhances Preservation of Normal but Not Chronic Myelogenous Leukemia Primitive Hematopoietic Progenitors. *Exp Hematol* 30, 324-332 (2002)

Schiedlmeier B, Schilz AJ, Kühlcke K, et al. Multidrug Resistance 1 Gene Transfer Can Confer Chemoprotection to Human Peripheral Blood Progenitor Cells Engrafted in Immunodeficient Mice. *Hum Gene Ther* 13, 233-242 (2002)

Kobune M, Xu Y, Baum C, et al. Retrovirus-Mediated Expression of the Base Excision Repair Proteins, Formamidopyrimidine DNA Glycosylase or Human Oxoguanine DNA Glycosylase, Protects Hematopoietic Cells from N,N',N''-Triethylenethiophosphoramidate (thioTEPA)-Induced Toxicity *In Vitro* and *In Vivo*. *Cancer Res* 61(13),5116-5125 (2001)

Kurre P, Morris J, Horn PA, et al. Gene Transfer into Baboon Repopulating Cells: A Comparison of FLT-3 Ligand and Megakaryocyte Growth and Development Factor Versus IL-3 During *Ex Vivo* Transduction. *Mol Ther* 3(6), 920-927 (2001)

Goerner M, Horn PA, Peterson L, et al. Sustained Multilineage Gene Persistence and Expression in Dogs Transplanted with CD34⁺ Marrow Cells Transduced by RD114-Pseudotype Oncoretrovirus Vectors. *Blood* 98(7), 2065-2070 (2001)

Kaneko S, Onodera M, Fujiki Y, et al. Simplified Retroviral Vector GCsap with Murine Stem Cell Virus Long Terminal Repeat Allows High and Continued Expression of Enhanced Green Fluorescent Protein by Human Hematopoietic Progenitors Engrafted in Nonobese Diabetic/Severe Combined Immunodeficient Mice. *Hum Gene Ther* 12, 35-44 (2001)

A report by French physician-scientists suggests a successful application of gene transfer methods in the treatment of two children with severe combined immunodeficiency (SCID) due to defective interleukin 2 receptor common gamma chain. The protocol used in this clinical trial was derived from a number of preclinical and basic studies leading to improved transduction of Hematopoietic stem and primitive progenitor cells using retrovirus vectors. These improvements have also been shown to impact transduction of a long-lived progenitor cell in a chemotherapy protocol in cancer patients. The improved results of these human trials come during a

period of increased scrutiny and criticism of human gene therapy trials, due, in part, to significant toxicities in some trials using adenovirus-based vectors. The potential efficacy versus toxicity of phase I trials of human gene therapy is also under question. After many years of research, however, there appears to be real evidence that genetic diseases may be successfully treated by gene transfer techniques. Future clinical studies should be based on continued progress in the understanding of the toxicology of gene delivery systems, vector technology, and target cell manipulation.

[Williams DA et al., review 2000]

- Bajaj B, Lei P, Andreadis ST. High Efficiencies of Gene Transfer with Immobilized Recombinant Retrovirus Kinetics and Optimization. *Biotechnol Prog* 17(4), 587-596 (2001)
- Pollok K, Van Der Loo JCM, Cooper RJ, et al. Differential Transduction Efficiency of SCID-Repopulating Cells Derived from Umbilical Cord Blood and Granulocyte Colony-Stimulating Factor-Mobilized Peripheral Blood. *Hum Gene Ther* 12, 2095-2108 (2001)
- Kapur R, Cooper R, Zhang L, et al. Cross-Talk between $\alpha_4\beta_1/\alpha_5\beta_1$ and c-Kit Results in Opposing Effect on Growth and Survival of Hematopoietic Cells via the Activation of Focal Adhesion Kinase, Mitogen-Activated Protein Kinase, and Akt Signaling Pathways. *Blood* 97(7), 1975-1981 (2001)
- Donahue RE, Dunbar CE. Update on the Use of Nonhuman Primate Models for Preclinical Testing of Gene Therapy Approaches Targeting Hematopoietic Cells. *Hum Gene Ther* 12, 607-617 (2001)
- Hacein-bey S, Gross F, Nusbaum P, et al. Optimization of Retroviral Gene Transfer Protocol to Maintain the Lymphoid Potential of Progenitor Cells. *Hum Gene Ther* 12, 291-301 (2001)
- Takatoku M, Sellers S, Agricola BA, et al. Avoidance of Stimulation Improves Engraftment of Cultured and Retrovirally Transduced Hematopoietic Cells in Primates. *J Clin Invest* 108(3), 447-455 (2001)
- Donahue RE, Wersto RP, Allay JA, et al. High Levels of Lymphoid Expression of Enhanced Green Fluorescent Protein in Nonhuman Primates Transplanted with Cytokine-Mobilized Peripheral Blood CD34(+) Cells. *Blood* 95(2), 445-452 (2000)
- Barquinero J, Segovia JC, Ramirez M, et al. Efficient Transduction of Human Hematopoietic Repopulating Cells Generating Stable Engraftment of Transgene-Expressing Cells in NOD/SCID Mice. *Blood* 95(10), 3085-3093 (2000)
- Kelly PF, Vandergriff JV, Nathwani A, et al. Highly Efficient Gene Transfer into Human Cord Blood NOD/SCID Repopulating Cells by Oncoretroviral Vector Particles Pseudotyped with the Feline Endogeneous Retrovirus (RD114) Envelope Protein. *Blood* 96(4), 1206-1214 (2000)
- Haas DL, Case SS, Crooks GM, Kohn DB. Critical Factors Influencing Stable Transduction of Human CD34(+) Cells with HIV-1-Derived Lentiviral Vectors. *Mol Ther* 2(1), 71-80 (2000)
- Goerner M, Roecklein B, Totok-Storb B, et al. Expansion and Transduction of Nonenriched Human Cord Blood Cells Using HS-5 Conditioned Medium and FLT3-L. *J Hematother Stem Cell Res* 9(5), 759-765 (2000)
- Shields LE, Kiem HP and Andrews RG. Highly Efficient Gene Transfer into Preterm CD34 Hematopoietic Progenitor Cells. *Am J Obstet Gynecol* 183(3), 732-737 (2000)
- Kim HJ, Tisdale JF, Wu T, et al. Many Multipotential Gene-Marked Progenitor or Stem Cell Clones Contribute to Hematopoiesis in Nonhuman Primates. *Blood* 86(1), 1-8 (2000)
- Wu T, Kim HJ, Sellers SE, et al. Prolonged High-Level Detection of Retrovirally Marked Hematopoietic Cells in Nonhuman Primates After Transduction of CD34+ Progenitors Using Clinically Feasible Methods. *Mol Ther* 1(3), 285-293 (2000)
- Wognum AW, Visser TP, Peters K, et al. Stimulation of Mouse Bone Marrow Cells with Kit Ligand, FLT3 Ligand, and Thrombopoietin Leads to Efficient Retrovirus-Mediated Gene Transfer to Stem Cells, Whereas Interleukin 3 and Interleukin 11 Reduce Transduction of Short- and Long-Term Repopulating Cells. *Hum Gene Ther* 11, 2129-2141 (2000)
- Trarbach T, Greifenberg S, Bardenheuer W, et al. Optimized Retroviral Transduction Protocol for Human Progenitor Cells Utilizing Fibronectin Fragments. *Cytherapy* 2(6), 429-438 (2000)
- Kume A, Xu R, Ueda Y, et al. Long-Term Tracking of Murine Hematopoietic Cells Transduced with a Bicistronic Retrovirus Containing CD24 and EGFP Genes. *Gene Ther* 7, 1193-1199 (2000)
- Verhasselt B, Naessens E, Smedt MD, et al. Efficiency of Transgenic T Cell Generation from Gene-Marked Cultured Human CD34+ Cord Blood Cells is Determined by Their Maturity and the Cytokines Present in the Culture Medium. *Gene Ther* 7, 830-836 (2000)
- Wilcox DA, Olsen JC, Ishizawa L, et al. Megakaryocyte-Targeted Synthesis of the Integrin β_3 -Subunit Results in the Phenotypic Correction of Glanzmann Thrombasthenia. *Blood* 95(12), 3645-3652 (2000)
- Emery DW, Andrews RG and Papayannopoulou T. Differences Among Nonhuman Primates in Susceptibility to Bone Marrow Progenitor Transduction with Retrovirus Vectors. *Gene Ther* 7, 359-367 (2000)

As the ultimate goal is the achievement of successful gene therapy, it is essential to transfer specific genes into target cells at high efficiency, and to maintain long-term stable expression by optimizing transduction conditions. Ongoing optimization efforts are focused on establishing better transfer systems by modifying oncoretroviral vector designs and pseudotyping with alternative envelope proteins, and by incorporating safety-modified HIV-1 based lentiviral vectors, together with further understanding of the biology of hematopoietic cells. The development of simple and clinically applicable fibronectin (FN CH-296)-assisted protocols that obviate the need for cocultivation and extended *ex vivo* manipulations with multiple exposures to viruses has been improving gene therapy trials and will make them more successful.

Chono H, Yoshioka H, Ueno M, et al. Removal of Inhibitory Substances with Recombinant Fibronectin-CH-296 Plates Enhances the Retroviral Transduction Efficiency of CD34⁺CD38⁻ Bone Marrow Cells. *J Biochem* 130, 331-334 (2001)

Demaison C, Brouns G, Blundell MP, et al. A Defined Window for Efficient Gene Marking of Severe Combined Immunodeficient-Repopulating Cells Using a Gibbon Ape Leukemia Virus-Pseudotyped Retroviral Vector. *Hum Gene Ther* 11, 91-100 (2000)

Garcia-Ortiz MJ, Serrano F, Abad JL, et al. Δ hGHR, a Novel Biosafe Cell Surface -Labeling Molecule for Analysis and Selection of Genetically Transduced Human Cells. *Hum Gene Ther* 11, 333-346 (2000)

Von Laer D, Corovic A, Vogt B, et al. Amphotropic and VSV-G-Pseudotyped Retroviral Vectors Transduce Human Hematopoietic Progenitor Cells with Similar Efficiency. *Bone Marrow Transplant* 26, Suppl 2, S75-S79 (2000)

Williams DA. Retroviral-Fibronectin Interactions in Transduction of Mammalian Cells. *Ann NY Acad Sci* 30(872), 109-113 (1999)

Gene transfer into haematopoietic stem cells has become an important strategy to tackle a number of inherited disorders of blood cell and immune system development. It can also be used in the treatment of leukaemia and other cancers if drug resistance genes are inserted into non-malignant stem cells before the tumour cells are attacked with chemotherapeutics (Abonour et al., 2000).

High-rate virus production in specific cell lines and pseudotyping these viruses with foreign proteins, such as Gibbon ape leukaemia virus and RD114 feline retrovirus envelope, significantly increase the ability of these vectors to specifically transfect their target cells. Researchers have modified retroviral vectors to enhance expression and avoid silencing of the gene after its insertion in the target cell's genome. Another important finding is that fibronectin and CH-296, a recombinant fragment of this pro-

tein, can significantly improve the efficiency of retroviral gene transfer. In addition, a better understanding of how growth factors regulate the development of blood and immune cells enables researchers to direct their vectors to the right stem cells. For instance, it has recently been shown that a combined use of various cytokines can induce the proliferation of human CD34 (+) CD38 (-) cells. The pretreatment makes these primitive haematopoietic cells permissive for integration of onco-retroviral provirus into their genome without inducing further differentiation. Stem cells with the therapeutic gene do not differentiate and thus get lost, but produce transduced daughter cells for a long time.

Although they come with limitations, these findings have provided sufficient results in gene therapy research to make clinical applications possible in selected settings.

[Fischer A, review 2000]

Murray L, Luens K, Tushinski R, et al. Optimization of Retroviral Gene Transduction of Mobilized Primitive Hematopoietic Progenitors by Using Thrombopoietin, Flt3, and Kit Ligands and RetroNectin Culture. *Hum Gene Ther* 10, 1743-1752 (1999)

Garrett E, Grain MI, Miller ARM, et al. Enhanced Retroviral Gene Transfer into CML and Normal Bone Marrow, and CML and Mobilized Peripheral Blood CD34⁺ Cells Using the Recombinant Fibronectin Fragment CH-296. *Br J Haematol* 107, 401-408 (1999)

MacNeill EC, Hanenberg H, Pollok KE, et al. Simultaneous Infection with Retroviruses Pseudotyped with Different Envelope Proteins Bypasses Viral Receptor Interference Associated with Colocalization of gs70 and Target Cells on Fibronectin CH-296. *J Virol* 73(5), 3960-3967 (1999)

Rebel VI, Tanaka M, Lee J-S, et al. One-Day *Ex Vivo* Culture Allows Effective Gene Transfer into Human Nonobese Diabetic/Severe Combined Immune-Deficient Repopulating Cells Using High-Titer Vesicular Stomatitis Virus G Protein Pseudotyped Retrovirus. *Blood* 93(7), 2217-2224 (1999)

Case SS, Price MA, Jordan CT, et al. Stable Transduction of Quiescent CD34⁺CD38⁻ Human Hematopoietic Cells by HIV-1-Based Lentiviral Vectors. *Proc Natl Acad Sci USA* 96, 2988-2993 (1999)

Krämer A, Hörner S, Willer A, et al. Adhesion to Fibronectin Stimulates Proliferation of Wild-Type and *bcr/abl*-Transfected Murine Hematopoietic Cells. *Proc Natl Acad Sci USA* 96, 2087-2092 (1999)

Dao MA, Taylor N, Nolte JA. Reduction in Levels of the Cyclin-Dependent Kinase Inhibitor p27^{kip-1} Coupled with Transforming Growth Factor β Neutralization Induces Cell-Cycle Entry and Increases Retroviral Transduction of Primitive Human Hematopoietic Cells. *Proc Natl Acad Sci USA* 95, 13006-13011 (1998)

Dao MA, Hashino K, Kato I, et al. Adhesion to Fibronectin Maintains Regenerative Capacity During *Ex Vivo* Culture and Transduction of Human Hematopoietic Stem and Progenitor Cells. *Blood* 92(12), 4612-4621 (1998)

van der Loo JCM, Xiao X, McMillin D, et al. VLA-5 Is Expressed by Mouse and Human Long-term Repopulating Hematopoietic Cells and Mediates Adhesion to Extracellular Matrix Protein Fibronectin. *J Clin Invest* 102(5), 1051-1061 (1998)

Pollok KE, Hanenberg H, Noblitt TW, et al. High-Efficiency Gene Transfer into Normal and Adenosine Deaminase-Deficient T Lymphocytes Is Mediated by Transduction on Recombinant Fibronectin Fragments. *J Virol* 72(6), 4882-4892 (1998)

Movassagh M, et al. High-Level Gene Transfer to Cord Blood Progenitors Using Gibbon Ape Leukemia Virus Pseudotype Retroviral Vectors and an Improved Clinically Applicable Protocol. *Hum Gene Ther* 9(2), 225-234 (1998)

Veena P, et al. Delayed Targeting of Cytokine-Nonresponsive Human Bone Marrow CD34⁺ Cells With Retrovirus-Mediated Gene Transfer Enhances Transduction Efficiency and Long-Term Expression of Transduced Genes. *Blood* 91(10), 3693-3701 (1998)

Yokota T, et al. Growth-Supporting Activities of Fibronectin on Hematopoietic Stem/Progenitor Cells *In Vitro* and *In Vivo*: Structural Requirement for Fibronectin Activities of CS1 and Cell-Binding Domains. *Blood* 91(9), 3263-3272 (1998)

Hanenberg H, Hashino K, Konishi H, et al. Optimization of Fibronectin-Assisted Retroviral Gene Transfer into Human CD34⁺ Hematopoietic Cells. *Hum Gene Ther* 8(18), 2193-2206 (1997)

Note : Shown above are a limited number of citations among those that have been published since 1996 reporting results obtained from studies using FN CH-296 or RetroNectinTM. Most articles published in the 1990's are not included, except those of seminal nature, due to space constraints. *SYP, Aug 2002*



TAKARA BIO INC.

The Biotechnology Company™

Otsu, Shiga, Japan Phone:+81 77-543-7247 Fax:+81 77-543-9254
Homepage: <http://www.takara-bio.co.jp> E-mail: bio-sm@takara-bio.co.jp

Takara Korea Biomedical Inc. Phone: (02) 577-2002 Fax: (02) 577-3691
TaKaRa Biotechnology (Dalian) Co., Ltd. Phone: (0411) 763-2792 Fax: (0411) 761-9946
TaKaRa Biomedical Europe S.A. Phone: +33 1 41 47 23 70 Fax: +33 1 41 47 23 71